



**ResourcePath, LLC**  
45945 Trefoil Ln #175  
Sterling, VA 20166  
571-375-0755  
Medical Director: D. Ashley Hill, MD  
hill@resourcepath.net

## **DICER1 Mutation Analysis Specimen Requirements and Collection Instructions**

### **We accept the following specimen types for DICER1 testing:**

- ✓ Whole Blood in EDTA (purple top) or PAXGene Tube
- ✓ Extracted DNA
- ✓ Saliva
- ✓ Buccal Swabs
- ✓ FFPE blocks
- ✓ FFPE scrolls
- ✓ Unstained Slides (less preferred)

- **PLEASE NOTE:** All specimen containers *must* be labeled with the patient's first and last name, date of birth, date and time of collection. Please include any other identifying information that you would like included in the report on the requisition form. *For FFPE scrolls of tumor tissue, please include an estimated percent tumor content for specimens sent on the requisition form.*
- **For each patient, please complete and send back the DICER1 Testing Requisition Form and the Clinical Consent for Genetic Testing paperwork.**
- The lab accepts specimens Monday through Friday. For samples collected on a Friday, please place them at refrigerated conditions and ship the following Monday using the standard overnight shipping option.
- Please ensure that all specimen collection supplies are used within their expiration date.
- All materials must be properly packaged and labeled to indicate the general nature of the materials being transported. The laboratory only accepts specimens considered Class B/non-infectious, and should be shipped using a FedEx Clinical Pak or equivalent.
- Please ship all specimens to:  
ResourcePath, LLC  
Attn: Accessioning  
45945, Trefoil Ln, Unit 175  
Sterling, VA 20166  
Phone for delivery questions: 314-313-8212

Please send an email to [info@resourcepath.net](mailto:info@resourcepath.net) when the specimens have shipped and include the following information:

- Date shipped
- Carrier
- Tracking number
- Number of specimens



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### **Whole Blood**

**Patient Preparation:** No special patient preparations are required

**Specimen Collection Container:** Either purple top/EDTA vacutainer or PaxGene Blood DNA tube

**Labeling:** All tubes must be labeled with the patient's first and last name, birthdate, date and time of collection.

**Procedure:**

1. Collect 4mls of blood into a purple top/EDTA tube OR 8mLs of blood into a PaxGene Blood DNA tube.

**Shipping:** Ship samples and paperwork overnight at room temperature. In hot climates, ship the samples in a package containing an ice pack. Place specimen into a plastic or padded container to protect the tubes from breaking.

### **Extracted DNA**

**NOTE:** DNA samples that are submitted for testing must be extracted in a CLIA certified laboratory. Please indicate CLIA license number on the requisition when submitting a DNA specimen.

**Patient Preparation:** Follow the patient preparation instructions based on the parent sample the DNA is being extracted from.

**Specimen Collection Container:** DNA samples can be placed in a 1.5ul sterile Eppendorf microcentrifuge tube or equivalent. Please implement good laboratory practice procedures for aliquotting DNA samples.

**Labeling:** All tubes containing DNA must be labeled with the patient's first and last name, date of birth, date and time of extraction. If the DNA specimen is extracted from tumor, please include the percent tumor content observed in the parent specimen.

**Procedure:**

1. Please send DNA that has been extracted by a method that has been verified by the literature or a commercially available kit.
2. Minimum sample requirements for **germline (non-fixed tissue)** DNA are as follows:
  - 5-10ug of DNA; minimum of 2ug.
  - Concentration: 100ng/ul
  - 260/280 ratio: 1.7
3. Minimum sample requirements for **tumor (fixed and fresh)** DNA are as follows:
  - 1ug of DNA; minimum 500ng
  - Concentration: 20ng/ul
  - 260/280 ratio: 1.7
4. Please label the tubes appropriately and secure the lids to ensure that the containers do not open during shipment.

**Shipping:** Ship samples and paperwork overnight on a cold pack. Please package the tubes to ensure that they will not sustain damage during transit.

## **DICER1 Mutation Analysis Specimen Requirements and Collection Instructions**

### **Saliva**

**Patient Preparation:** Please ensure the patient has not had anything to eat or drink 30 minutes prior to collection.

**Specimen Collection Container:** IBI Scientific Saliva Collection and Prep Kit

**Labeling:** All tubes must be labeled with the patient's first and last name, birthdate, date and time of collection.

**Procedure:**

1. Remove the snap-on cap from the collection funnel.
2. Spit into the center of the funnel and fill the collection tube to the solid line that goes all the way around the tube (this is the 2ml mark).
3. Remove the green cap from the 5ml saliva suspension solution tube. Pour the entire contents of the saliva suspension solution into the collection funnel.
4. Remove the collection funnel from the saliva storage tube and discard.
5. Place the accessory screw cap on the saliva storage tube and tighten. Shake sample tube well to properly mix the saliva with the suspension solution.

**Shipping:** Ship samples and paperwork overnight at room temperature. Please package samples securely in a padded envelope or a plastic container to protect the tubes from breaking in transit

### **Buccal Swabs**

**Patient Preparation:** Please ensure the patient has not had anything to eat or drink 30 minutes prior to collection.

**Specimen Collection Container:** IBI Scientific Saliva Collection and Prep Kit and flocked swabs for buccal cell collection

**Labeling:** All containers must be labeled with the patient's first and last name, date of birth, date and time of collection.

**Procedure:**

1. Verify that the subject's mouth is empty.
2. Wash or sanitize hands.
3. Open the snap-on cap on the saliva collection funnel attached to the saliva collection tube.
4. Carefully remove swab from package. Avoid touching the tip of the swab to any surfaces.
5. Gently rub the swab on the inside of the subject's cheek for 5-10 seconds, ensuring that the entire swab has contacted the cheek.
6. Remove the swab, being careful not to touch it to the subject's teeth or any other surfaces.
7. Break or cut the swab stick so that the swab will fit in the saliva collection tube.
8. Repeat steps 3-6 for the second swab.
6. Once both swabs have been collected, remove the green cap from the 5ml saliva suspension solution tube. Pour the entire contents of the saliva suspension solution into the collection funnel.
7. Remove the collection funnel from the saliva storage tube and discard.
8. Place the accessory screw cap on the saliva storage tube and tighten. Shake sample tube well to properly mix the saliva with the suspension solution.

**Shipping:** Ship samples and paperwork overnight at room temperature. Please package samples securely in a padded envelope or a plastic container to protect the tubes from breaking in transit.



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### **FFPE Blocks**

**Patient Preparation:** Please ensure that there is tissue visible in the blocks that are sent.

**Specimen Collection Container:** Your laboratory's preferred tissue cartridge

**Labeling:** All blocks must be labeled with the patient's first and last name, date of birth, date and time of collection, and percent tumor content.

**Procedure:**

1. Prepare fixed tissue per your laboratory's procedure.

**Shipping:** Please package the blocks to ensure they will not sustain damage during transit. It is not necessary to ship these samples overnight but please remember that our turnaround time guarantee is from the receipt of specimen.

### **FFPE Scrolls**

**Patient Preparation:** Please ensure that there is tissue visible in the blocks that scrolls are cut from.

**Specimen Collection Container:** Please place scrolls in a sterile container such as a 1.5ul Eppendorf microcentrifuge tube or equivalent.

**Labeling:** All tubes containing scrolls must be labeled with the patient's first and last name, date of birth, date and time of collection, and percent tumor content observed in the parent block.

**Procedure:**

1. Clean microtome blades and surface with 70% ethanol and allow to dry before cutting tissue.
2. Cut 2-10, 5um scrolls and place them in sterile container(s), being careful not to allow the scrolls to touch any surfaces that have not been decontaminated.
3. Label each tube appropriately and secure the top with parafilm to ensure that the containers do not open during shipment.

**Shipping:** Please package the tubes to ensure that they will not sustain damage during transit. It is not necessary to ship these samples overnight but please remember that our turnaround time guarantee is from the receipt of specimen.

### **Unstained Slides**

**Patient Preparation:** Please ensure that the slides have not been stained.

**Specimen Collection Container:** Glass slides.

**Labeling:** All slides must be labeled with the patient's first and last name, date of birth, date and time of collection, and percent tumor content.

**Procedure:**

1. Prepare slides per your laboratory's procedure for preparing unstained slides.

**Shipping:** Please package slides in a plastic slide cartridge and secure in a padded envelope to protect the slides from breaking in transit. It is not necessary to ship these samples overnight but please remember that our turnaround time guarantee is from the receipt of specimen.